Alveolar sampling and fast kinetics of tetrachloroethene in man. II Fast kinetics

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ABSTRACT Fast kinetic phenomena were studied in human subjects exposed to tetrachloroethene (perchloroethylene, PER). The duration of exposure ranged from one to 60 minutes and the concentration of PER in inhaled air ranged from 0.02 to 0.40 mmol/m³. The PER concentration in mixed venous blood (pulmonary artery) was estimated by alveolar concentration (C_{Alv}) measured after a residence time of 10 s. During exposure, stoppage of intake (breath holding up to 50 s) caused a decrease of C_{Alv} down to about 60% of the C_{Alv} at a residence time of 10 s. At the end of exposure, stoppage of intake (breathing fresh air) caused a decrease of C_{Alv} with $t_{\frac{1}{2}} = 15-25$ s; after two to four minutes, the decrease slowed down abruptly and the concentration remained more or less constant for about one to three minutes. After this stationary level, the decrease of C_{Alv} continued but at a slower rate. During and after exposure, the decrease of C_{Alv} seems to be caused by large differences in the circulation times of blood flowing through rapid, well perfused tissues and slower, well perfused tissues which may explain the stationary level. From this point of view, the vessel rich group in a compartment model must be split up in order to predict tissue and organ concentrations during peak environmental concentrations.

Human biological monitoring is a valuable tool with which to estimate quantitatively exposure to toxic agents. The exposure level is usually determined as a time weighted average; this means that peak concentrations are seldom taken into account.

Rapid changes in environmental concentrations may result in rapid and large differences in the concentration in organs or tissues. The distribution of a toxic agent in the human body is often described by compartment models in which the blood circulation times are set at zero. In the models the concentration of the agent in several groups of tissues and organs is often calculated by means of the estimated tissue volume, blood perfusion, and the tissue blood partition coefficient.¹

A more detailed modelling may be useful in predicting the magnitude and duration of differences in concentration in organs and tissues during and after peak concentration.

Mapleson suggested that for a few minutes after a change of the inhaled concentration the various circulation times of blood through the tissues and organs might play an important part in the time course of the concentration in mixed venous blood.² The concen-

tration in mixed venous blood (C_{Ven}) presents a mixture of blood coming from local organs and tissues with local perfusion, partition coefficients, and circulation times.

Frequent sampling of mixed venous blood after a change in the inhaled concentration is almost impossible but the sampling of alveolar air provides a valid estimate of the mixed venous concentration and makes it possible instantaneously to follow fast changes in C_{ven}. From exposure tests with tetrachloroethene (perchloroethylene, PER), we have shown that alveolar sampling in the postexposure period had much less variability than during exposure.³

The purpose of the present study was to assess fast kinetic phenomena in the early postexposure time after the abrupt cessation of respiratory intake of PER.

Methods

PROCEDURE

Six volunteers (3 men, 3 women) were exposed several times at rest to constant concentrations of PER in air.

The exposures, apparatus, sampling, sampling errors, and analysis have been described elsewhere.³

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In that paper the alveolar sampling technique was studied by taking samples after several episodes of breath holding. A distinction was made between breath holding time and residence time (t*): the latter was defined as the time interval between the beginning of inhalation and the end of the next exhalation. All samples taken after a certain breath holding time are expressed with their corresponding residence times. At a respiratory frequency of 12/0, a zero breath holding time corresponds to $t^* \simeq 5$ s.

In the early postexposure period (0-30 min) an alveolar sample was sometimes taken from each second or third exhalation after a 5, 10, or 20 s residence time. Extra attention was also paid to long breath holding—that is, t* > 10 s—during exposure.

Results

COURSE OF ALVEOLAR CONCENTRATION

Figure 1 shows an overall view of the course of the alveolar concentration (C_{Alv}). It has been shown that during exposure C_{Alv} decreases with increasing resi-

dence time.³ At $t^* \le 10$ s, the decrease of C_{Alv} may be described by an exponential curve; at about $t^* = 10$ s. the curve tends to stabilise at about 80% of C_{Alv} (t* = 5 s). At $t^* > 10$ s, however, C_{Alv} decreases more rapidly again; this decrease continues up to at least t* = 55 s, and the alveolar concentration is about 40% of the alveolar value at $t^* = 5$ s.

Immediately after the end of exposure, CAIN decreases rapidly. After a short period (t_2-t_1) of two to four minutes, the rate of decrease slows down abruptly and drops almost to zero for a short period $(t_3-t_2).$

In the postexposure period at $t-t_1 > 0.5$ min, the $(t^* = 5 s)$ values are lower than the $(t^* = 10 s)$ values and they both decrease in a constant ratio of 0.93 (table 1).

In the first 3.5 minutes of the postexposure period, the ($t^* = 10$ s) values are lower than the ($t^* = 20$ s) values; both decrease in a ratio of 0.90. During the next 30 minutes, this ratio becomes 0.96 whereas after four hours the ($t^* = 10 \text{ s}$) and ($t^* = 20 \text{ s}$) values are no longer distinguishable (table 1).

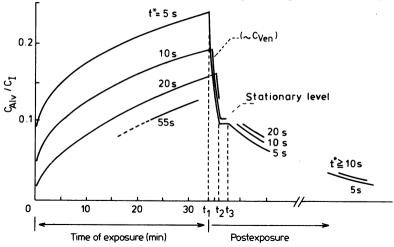


Fig 1 Overall view of alveolar concentration curves relative to inhaled concentration (C_1) of PER at different residence times (t^*) . In postexposure period lines are drawn schematically. At t > 240 min there was no difference between sampling with $t^* = 10$ s and $t^* \ge 20$ s.

Table 1 Difference in alveolar concentration of tetrachloroethene (PER) at different residence times in the postexposure period*

	$\frac{C_{Alv}(t^* = 5 s)}{C_{Alv}(t^* = 10 s)}$	$\frac{C_{Alv}(t^* = 10 \text{ s})}{C_{Alv}(t^* = 20 \text{ s})}$	No of	
Postexposure time (min)			Data	Experiments
0·5–4000 1·5–3·5† 4–35‡ 4–1500	0·93 ± 0·04	$\begin{array}{c} 0.90 \pm 0.02 \\ 0.96 \pm 0.02 \\ 0.98 \pm 0.02 \end{array}$	14 4 9 15	5 4 6 6

^{*}Values expressed as ratios ($\bar{x} \pm SD$). †Short postexposure period before stationary level. ‡Postexposure period beyond stationary level.

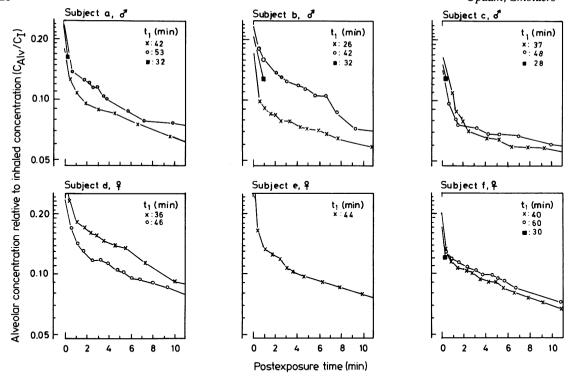


Fig 2 Semilog plots of C_{Alv}/C_1 in early postexposure period for six subjects. Different experiments with same subject are represented by symbols \bigcirc , \times , and \blacksquare ; symbol \blacksquare was measured from first exhalation after a short period of breath holding immediately after exposure at t_1 ; t_1 is duration of exposure. Extrapolation to zero postexposure time points in direction of C_{Alv} ($t^* = 10 \text{ s}$) at t_1 .

STATIONARY LEVEL

Figure 2 shows semilog plots of the time courses of C_{AIv} (t* = 10 s) in the early postexposure period for six subjects. In most experiments a fast rate of decrease is followed abruptly by a much smaller rate for a short period. During this short period, C_{AIv} sometimes tends to remain at a constant level: a stationary level. This short period ranged from about one (subjects a, e) to three minutes.

Beyond the stationary level the rate of decrease increases again and yields a smooth exponential decrease. Two exponential curve fitting procedures were carried out with the experimental data beyond the stationary level up to 60 minutes after exposure (appendix 2, eq 2); the curve could be fitted with a weighted residual error of only 2.5% (appendix 2, eq 3). Because of the stationary level it is difficult to fit a smooth multiexponential decrease for the whole postexposure period. The results in fig 2 do not, in general show a clear constant concentration level. For a short period, however, the concentrations seem more or less constant (a level); beyond this level the rate of CAIV decrease increases again. Because of the small weighted residual error around the fitted curve with the data beyond the stationary level, this level and the increase of the rate of C_{Alv} decrease cannot be explained by random or systematical errors.

There was usually some intraindividual variability in the course of C_{Alv} during exposure between the experiments. In figure 2 this becomes obvious from the C_{Alv} value at zero post exposure time (subjects a, c, and d). The postexposure courses of C_{Alv} of subject d may be explained by hyperventilation during the 36 minute exposure (symbol \times). Its total uptake exceeded the total uptake during the 48 minute exposure (symbol \bigcirc).

Figure 3 presents both the relative alveolar concentration (C_{Alv} (t* = 10 s)) at the end of exposure and the concentration of the stationary level (C_{SL}) as functions of the duration of exposure in 17 experiments on subject a.

FAST POSTEXPOSURE DECREASE OF ALVEOLAR CONCENTRATION

The actual time of exposure ends when the volume of the last inhalation equilibrates with C_{Ven} —that is, the net gas transfer into blood is zero. At this time (t_1) the alveolar concentration has the value of C_{Alv} $(t^* = 10 \text{ s})$ and decreases at $t > t_1$ because the uptake of PER has stopped.

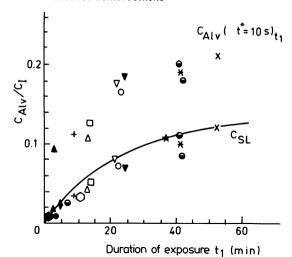


Fig 3 Relative alveolar concentration of PER at end of exposure (C_{Alv} ($t^* = 10 \text{ s}$) t_1 : upper symbols) and corresponding relative stationary level concentration in experiments on subject a; each experiment has same symbols. Drawn line was calculated by means of equation 1 (appendix 2) with estimated two coefficients and two exponents which were obtained by two exponential curve fitting (eq 2, appendix 2) with measured concentrations beyond stationary level ($t > t_3$) of one experiment (symbol Θ).

The experiments showed a multiexponential decrease of C_{AIv} (t* = 10 s), which is representative of mixed venous blood; it starts with C_{AIv} (t* = 10 s) at t_1 and approaches the stationary level C_{SL} at t_2 . The subjects showed for $t_1 < t < t_2$ an intra/intervariability in the rate of C_{AIv} decrease with t_2 = 15–25 s at t_1 , whereas after about 25 s the t_2 ranged from 20 to 50 s.

The $t_{\frac{1}{2}}$ is defined as the halftime of the C_{Alv} decrease between C_{Alv} (t* = 10 s) at t_1 and C_{SL} at t_2 . The half times at the end of exposure were obtained from a semilog plot of the data. These data were obtained from frequent sampling—for example from each second or third exhalation—and fitted by hand.

No relation was observed between duration of exposure (t_1) and half time $(t_{\frac{1}{2}})$. In some experiments immediately after exposure the subject did not inhale fresh air but held his breath for 15–60 s. When compared with normal fresh air breathing, this breath holding does not substantially effect the $t_{\frac{1}{2}}$ value (fig 2, symbol \blacksquare).

Discussion

 $\label{eq:fast_calling} \textbf{Fast} \ \textbf{Calv-Decrease} \ \ \textbf{and} \ \ \textbf{The} \ \ \textbf{Stationary} \quad \cdots \\ \textbf{Level}$

 C_{Alv} equilibrates with C_{Ven} at $t^* = 10$ s; indicates that at $t^* > 10$ s, C_{Alv} represents C_{Ven} .³ Alveolar air sam-

pling was carried out discontinuously but frequent sampling with $t^* = 10$ s gives the rate of change in the mixed venous concentration. At the end of exposure. C_{Alv} (t* = 10 s) decreased rapidly until it reached a stationary level (fig 1). Breath holding at the end of exposure did not substantially delay the decrease. Thus the decrease of CAIV is not caused only by the respiratory excretion but mainly by a fast "internal" decrease in C_{Ven}. This C_{Ven} is a composite of venous concentrations coming from rapid and slower perfused organs and tissues. Some explanations for the fast C_{Alv} or C_{Ven} decrease at t₁ or both, may be suggested: (a) absorption of PER in the airways. (b) absorption by lipid contents in the lung tissue, and (c)fast clearance of PER by organs and tissues. Absorption by the airways is unlikely as PER is almost insoluble in water

Significant lipid contents in lung tissue might continuously absorb PER from the alveolar air and blood in the capillary bed. In this way a significant contribution to the alveolar decrease is not likely because the rate of C_{Alv} decrease does not depend on the duration of exposure and in the early post-exposure period breath holding ($t^* > 10$ s) does cause an increase of C_{Alv} (table 1 and fig 1).

The clearance of PER from the blood by unsaturated perfused organs and tissues may be significant; at t = 30 min, the alveolar retention still equals about 75% during normal breathing ($t^* = 5$ s) (fig 1). It is not likely that those perfused tissues (splanchnic tissue, muscles, and fat, for example) are capable of causing the fast C_{Ven} decrease at t₁. The fast C_{ven} decrease at t₁ implies that the local venous concentration coming from at least one organ, tissue, or shunt must decrease rapidly. Therefore, one has to consider the distribution of the blood. Table 2 shows the values for the distribution of blood for four compartments in a standard man. The total blood volume has been shared between compartments in the same proportions as the cardiac output which leads to the same (total) circulation tissue for all compartments. This distribution is often used in conventional compartment modelling in which the circulation times are set at zero.1 A more likely distribution is listed in table 3: the blood volume in each tissue in the vessel rich group (VRG) is based on estimated local blood flows and transit times.² The remaining blood volume has been shared between the muscle group (MG) and fat group (FG) compartment in proportion to their flows. Simulation for PER was carried out with a four compartment model that was written for trichloroethene¹; in the present paper the model was used with zero metabolism because of the negligible metabolism of PER. Simulation during exposure for one hour for a standard man (table 2) showed that the partial presses in the muscle (P_{MG}) and fat group (P_{FG}) are about 35% and 0.7% of the solvent arterial pressure (PArt). This means that the local venous concentrations coming from both groups continue to increase during some postexposure period until PMG $= P_{Art}$ or $P_{FG} = P_{Art}$. In addition it is not expected that the contribution to Cyen from both groups would decrease within 40 s after stoppage of intake because of their circulation times which are > 40 s.² Even the local venous concentration coming from the splanchnic tissue does not decrease within its corresponding circulation time of about 55 s (table 3). Nevertheless, the splanchnic tissue belongs to the VRG and would be expected to have a much higher local venous partial pressure than P_{MG} and P_{FG} at time t₁. Therefore, splanchnic tissue probably also plays an important part in the course of Cven in the early postexposure period because of its relative high contribution to Cven.

Those tissues, organs, or shunts from which the local venous concentrations are able to react almost immediately at t₁ must have a short circulation time, however, and a rather small distribution volume.

WELL PERFUSED COMPARTMENT WITH A SHORT CIRCULATION TIME

Some questions may arise about the fast phenomena superimposed during exposure on the slower Cs. concentration course (fig 3). The above discussion gives good reasons to suppose the existence of rapidly perfused organs, tissues, or shunts that have: (a) a short circulation time of 10-15 s, (b) a rather small capacity for PER, and (c) an ability to raise substantially the venous concentration coming from slower parts up to the estimated mixed venous concentration.

Mapleson has reviewed the data on transit times. tissue volumes, and blood volumes for a standard man² (table 3). The transit time is the flow time from local artery to local vein; for an organ or tissue, this time may be estimated by dividing the corresponding blood volume by the blood flow. The highway circulation time refers to the flow time from left ventricle to local artery and from local vein to right atrium: this is estimated to be about 5.5 s. The pulmonary circulation time is estimated to be 7.5 s; this time is common to circulations coming from all organs and tis-

Table 2 Tissue and blood volumes, blood perfusions, and partition coefficients of PER for standard man at rest according to several authors*

	Compartments			
	VRG	MG	FG	Pulmonary
Tissue volume (l) ⁶	8.8	36.3	11.5	1.0
Blood volume (1)6	3.19	0.63	0.18	1.4
Blood perfusion (l/min)	5.20	1.02	0.29	
Partition co-efficient:				
bl/air ⁴			15	
tis/bl	2·58	3.58	146⁴	19
Average circulation time (s)†	37	37	37	

^{*}Standard man: body weight 70 kg, body surface area 1.8 m², and cardiac output 6.5 l/min.

Table 3 Preferred tissue volumes, blood volume, blood flows (O), transit and total circulation times for a standard man^{2*}

	Volume (l)	Blood volume§ (1)	Q (l/min)	Transit time (s)	Total circ time (s)¶
Lung parenchymal tissue VRG	0.5				
Kidneys	0.28	0.052	1.25	2.5	15.5
Adrenals	0.02	0.004	0.10	2.5	15.5
Thyroid	0.02	0.004	0.10	2.5	15-5
Heart	0.28	0.03	0.24	6·5–11	19-5-24
Brains	1.43	0-11	0.79	8.5	21.5
Other glands and organs	0.18		0.05	8.5	21.5
Splanchnic tissue	3.73	1.05	1.53	41	54
Total	5.94	2.08	4.06		
Average time†		"		31	44
MG					
Total	37-4	2.17	1.13		
Average time†				115	128
FG					
Total	12.8	0.58	0.30		
Average time†				115	128
Peripheral shunt‡		0.56	1.04	32.5	40

[†]Average circulation time equals blood volume divided by blood flow.

^{*}Standard man: 70 kg, aged 30-39, 1.85 m² body surface area. †Average circulation time equals blood volume divided by blood flow

 $[\]pm$ In this paper both fast and slow shunt are assumed to have Q = 0.52 l/min.

Blood volume estimated as blood flow × transit time.

Including the blood volume (0.83 l) in the pulmonary circulation and heart chambers.

[¶]Total circulation time equals sum of transit time, highway (5.5 s) and pulmonary time (7.5 s).

sues. The total circulation time is the sum of transit time, highway circulation, and pulmonary circulation time.

Although the average circulation time for the visceral compartment (vessel rich, VRG) is estimated to be 44 s, a blood flow of 1.45 l/min returns to the lungs within 15.5 s; for a blood flow of 2.55 l/min, the time is 21.5 s (table 3). Blood coming through the splanchnic tissue, however, returns after about 55 s. In respect to circulation times, the visceral compartment may be split up into these subgroups: VRG 1a (kidney, adrenals, and thyroid, etc), VRG 1b (heart, brains, and other small glands and organs), and VRG 2 (splanchnic tissue).

The peripheral shunt in table 3 will probably also be a composite of several shunts. Arteriovenous anastamoses in the skin and muscles provide well known shunts. The average circulation time equals about 39 s but some shunts probably have much shorter times. The numerical values of bloodflows and circulation times of various shunts are uncertain, but the total peripheral shunt of table 3 can probably be split up into rapid and slow shunts; we assume both have equal blood flow ($\simeq 0.5 \text{ l/min}$) with circulation times of 10 s and 60 s respectively.

Figure 4 presents a compartment model; in view of

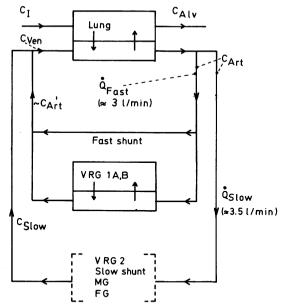


Fig 4 Compartment model based on physiological data of blood circulation times of organs and tissues which belong to vessel rich group (table 4). VRG is split into VRG 1 and VRG 2. Composition is assumed to be: VRG 1: kidney, adrenals, thyroid, heart, brains, other small glands, and organs; VRG 2: splanchnic tissue. Total arteriovenous shunt is split into a fast and a slow part.

the circulation times the VRG has been split up and fast and slow shunts have been added.

With the exception of the brain, all tissues in the VRG I compartment have a small tissue volume, a small (tissue/blood) partition coefficient (table 2) and therefore a small capacity (distribution volume) to absorb PER. Shunts will have an even smaller capacity. These very rapidly perfused tissues and organs will equilibrate quickly with C_{Art} and their local venous blood will react almost immediately to changes in C_{Art}.

The rise in the local venous concentration at a level C_{SL}^* ($\simeq C_{SL}$; fig 3) up to the C_{Ven} value (C_{Alv} at ($t^*=10~s$) may be plausibly explained. To put it simply, a composition of bloodflows with C_{SL}^* and C_{Arr} must yield C_{Ven} . During exposure, local venous blood coming from the shunt and VRG 1 tissues has a concentration almost equal to C_{Arr} (fig 4). Venous blood coming from slower VRG 2, MG, and FG compartments and slow shunt is mixed and results in a course of the concentration (C_{Slow}) which is assumed to be the same as the time course of the concentration at the stationary level (C_{SL}^*) (fig 3). Mixed venous blood may be regarded as a composite of the two mentioned venous blood flows: this yields a C_{Ven} value that is about 20% lower than the C_{Ven} value ($t^*=10~s$) (fig 1)

Although this calculated C_{Ven} value is rather low, it is shown that shunts and VRG 1 tissues can increase venous concentration considerably up to the mixed venous level measured at $t^* = 10$ s.

Simulations for PER with the four compartment model for a standard man (table 2) yielded a fast smooth decrease with a half time of > 0.5 min immediately after the end of exposure. Even the introduction of an artificially fast (highly perfused) small compartment does not simulate a stationary level. The estimated average circulation times of the three peripheral compartments appear about to be equal (table 2).

A monoexponential C_{Ven} decrease between C_{Ven} (t_1) and C_{SL}^* (t_2) which is assumed to be caused only by internal PER redistribution, may be estimated by means of a mass balance equation which describes a fast system (fig 4). This fast system consists of VRG 1, fast shunt, and the corresponding blood volume including the blood in pulmonary circulation and heart chambers (table 3). The parenchymal lung tissue is excluded because its local venous blood is not supplied to the mixed venous blood but to the arterial blood (left atrium). The mass balance equation describes the fast PER transport into and out of the fast circulation system. During (t_2 - t_1) C_{Slow} is

assumed to be of the same value as C*_{SL} at t₂; this is justified by the slow change of C_{Slow} at t₁ due to the slow change of the local venous concentrations coming from the slower perfused organs and tissues. The mass balance equation is:

$$\begin{split} V_{\text{Fast}}\,C_{\text{Ven}} &= C_{\text{Slow}}\,\dot{Q}_{\text{Slow}}\,\Delta t - C_{\text{Art}}\,\dot{Q}_{\text{Slow}}\,\Delta t \\ \text{where } C_{\text{Art}} &= C_{\text{Ven}}, \text{ due to the assumed zero respiratory excretion and } C_{\text{Slow}} &\simeq C^*_{\text{SL}}. \text{ The solution is:} \\ C_{\text{Ven}}(t) &= C^*_{\text{SL}} + (C_{\text{Ven}}(t_1) - C^*_{\text{SL}}) \\ &= \exp\left(\left(-\frac{\dot{Q}_{\text{Slow}}}{V_{\text{Fast}}}\right)\!(t-t_1)\right) \quad (1) \end{split}$$

The distribution volume (V_{Fast}) is defind as the total volume of the fast system which is instantaneously in equilibrium with local venous concentration during rapid changes in C_{Art} . Two extreme conditions are considered during the rapid C_{Ven} decrease: (a) all VRG 1 tissues—for example, kidney and brain—are assumed to be completely in equilibrium with the local venous concentration as it leaves them; this yields $V_{Fast} = 6.7$ 1, with $\lambda_{tis/bl} = 2.5$ and (b) VRG 1 tissues are not assumed to be in equilibrium with local venous concentration; there is no desorption and local venous concentration equals C_{Art} , the volume of the bloodflow in the fast parts equals $V_{Fast} = 1.2$ 1 (table 3).

Using equation 1, with $Q_{Slow} = 3.5$ l/min, the first condition gives a half time of 80 s whereas the second condition gives $t_{\frac{1}{2}} = 14$ s. During the first 10–30 s after stoppage of the uptake, desorption out of the VRG 1 tissues is unlikely to cause a much longer half time than $t_{\frac{1}{2}} = 14$ s. Probably after some time the first condition starts to dominate. The desorption becomes more important and this may explain the observed change of $t_{\frac{1}{2}}$ from 15 to 25 s to 20 to 50 s at about $t-t_1 = 20$ s.

Respiratory excretion can shorten the half time because of an extra output out of the fast system. This respiratory output is low compared with the output yielded by the bloodflow \dot{Q}_{Slow} .

In the early postexposure period C_{Ven} is strongly affected by large differences in (a) partial pressures in the fast perfused organs and tissues and (b) in their corresponding circulation times. This becomes apparent with sampling after $t^* = 20$ s which yields a 10% higher C_{Alv} value than C_{Alv} ($t^* = 10$ s) (table 1 and fig 1). It seems plausible that a relatively high local venous concentration coming from fast perfused organs, tissues, or shunts reacts on changes in respiratory excretion within 10-20 s and is able to affect the C_{Ven} value. In the later postexposure period ($t-t_1$) > 35 min large differences have disappeared and the fast perfused tissues and organs are no longer able to

affect C_{Ven} significantly: $C_{Alv}(t^* = 10 \text{ s}) \simeq C_{Alv}(t^* \ge 10 \text{ s})$ (table 1, fig 1).

It may be concluded that the decrease of the alveolar concentration at t₁ is mainly caused by the mixing of local venous blood coming from a rapid VRG 1 compartment and a rapid shunt with local venous blood coming from the slower compartments.

STATIONARY LEVEL AND $\mathbf{C}_{\mathbf{Alv}}$ Course beyond the level

In the postexposure period slower, well perfused organs and tissues are responsible for the course beyond the stationary level ($t > t_3$). A two compartment model was fitted to the data of CAIv beyond the stationary level of an experiment (appendix 2, eq 2). The coefficients and exponents $(c(1), c(2), r_1, r_2)$ of the fitted function (appendix 2, eq 2) were used to calculate a course of C_{Alv} during exposure by means of equation 1 (appendix 2); this C_{Alv} course reflects the venous concentration C_{Slow} during exposure. Figure 3 shows that the value of the stationary level in different experiments equals this calculated venous concentration. This suggests that the mixture of local concentrations coming mainly from the slower well perfused organs and tissues can be measured at first at the stationary level.

The splanchnic tissue is well perfused (1.5 l/min) and much more saturated than the muscles and fat in the postexposure period (0–60 min). Therefore, the contribution from the splanchnic tissue, which includes the liver, to C_{ven} is rather large. It seems plausible that the splanchnic tissue is most responsible for the existence and magnitude of the stationary level and also for the change in C_{ven} in the post level period up to about 60 minutes. This post-exposure period seems to offer the opportunity to study the kinetics of the splanchnic tissue in more detail. In particular this postexposure period might be important for assessing individual metabolism in the liver.

In summary, fast kinetic phenomena were studied by means of abrupt stoppage of respiratory intake. This was realised by long breath holding during exposure or by breathing fresh air immediately at the end of exposure. During exposure, breath holding does decrease alveolar concentration considerably. At the end of exposure, the subject inhaled fresh air and a similar decrease of alveolar concentration occurred which is scarcely delayed by breath holding. It was concluded that the rapid decrease in the mixed venous concentration causes a rapid decrease in the alveolar concentration. This rapid mixed venous decrease could be explained by the rapid venous blood returning from well perfused small organs, tissues, or shunts with short circulation times (10-20 s). After exposure, the decrease of alveolar concentration was

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not smooth but it slowed down abruptly after a few minutes and a short stationary level of one to three minutes occurred. Beyond this level, the decrease was continued at a slower rate. Riley et al found a similar pattern in the rate of alveolar decrease after exposure to methylene chloride but could offer no physiological explanation. ¹⁰ In this paper we suggest that the stationary level was due to mixed venous blood coming from more slowly perfused organs and tissues, in particular the splanchnic tissue. For the assessment of individual metabolism in the liver, the alveolar course beyond the stationary level might be important.

To assess the kinetics of fast phenomena, we suggest that a small compartment or shunt, or both, with a blood circulation time of 10–15 s must be added to the conventional compartment model; the bloodflows which belong to the other compartments have longer circulation times (about 40–55 s).

Alveolar air sampling is non-invasive and allows frequent sampling. As a result we were able to distinguish the stationary level and to assess the fast kinetics of a solvent in the body. This method may provide information on individual differences in fast kinetic data

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Appendix 1

SYMBOLS AND ABBREVIATIONS USED IN THE TEXT AND APPENDIX 2

AND APPENDIX 2	2
t*	Residence time; total time in which gas exchange can take place between inhaled alveolar air and blood in the capillary bed of the
t ₁	lung (s) Half time of C_{Alv} decrease between C_{Alv} (t* = 10 s) at t_1 and C_{SL} at t_2 (s)
t_1	Point of time at the end of
t_2	exposure (min) Point of time at the beginning of the stationary level (min)
t_3	Point of time at the end of stationary level (min)
C_{I}	Concentration of PER in inhaled
C_{Alv}	air (μg/l) during exposure Concentration of PER in alveolar air (μg/l)
C _{Alv} /C _I	Concentration of PER in alveolar air relative to the concentration in inhaled air

$C_{Alv}(t^* = x s)$	Concentration of PER in alveolar
	air after a residence time of x sec-
C (+* 10°)	onds (µg/l)
$C_{Alv}(t^*=10s)_{t_1}$	Concentration of PER in alveolar
	air after a residence time of 10 sec-
	onds at the end of exposure t_1
C D	(μg/l)
Cven, Pven	Concentration or partial pressure
	of PER in mixed venous blood
C D	returning to the lungs (µg/l, Pa)
C _{Art} , P _{Art}	Concentration or partial pressure
	of PER in arterial blood leaving
	the lungs (µg/l, Pa)
C _{SL}	Concentration in alveolar air at
	the stationary level between t2 and
	$t_3 (\mu g/l)$
C* _{SL}	Concentration in mixed venous
	blood at the stationary level
	between t_2 and t_3 —that is C_{SL}/λ ;
	$(\mu g/l)$
C _{Slow}	Concentration in venous blood
	coming from VRG2, MG, and FG
	compartments and slow shunts
	$(\mu g/l)$
VRG 1 a	Rapidly well perfused organs and
	tissues with a short transit time of
	two to three seconds: kidneys,
	adrenals thyroid, etc
VRG 1 b	Rapidly well perfused organs and
	tissues with a short transit time of
	6-11 seconds; heart, brains, and
	small glands and organs
VRG 1	VRG 1 a + VRG 1 b
VRG 2	Well perfused organs and tissues
	with a transit time of about 40 sec-
	onds: splanchnic tissue
VRG	Vessel rich group compartment:
	VRG 1 a + VRG 1 b + VRG 2
MG	Muscle group compartment
FG	Fat group compartment
Q _{Slow}	Blood flow returning to the lungs
	coming from VRG2, MG, and FG
	compartments and slow shunts
	(1/min)
Q	Cardiac output (l/min)
V_{Fast}	Distribution volume of the fast
	system which is composed by
	URG1, fast shunt, and a corre-
_	sponding blood volunme (1)
λ	Blood/air partition coefficient

Appendix 2

 $\lambda_{tis/bl}$

Compartment models have been used often in pharmokinetics. The number of compartments are often made equal to the number of exponents that are

Tissue/blood partition coefficient

necessary to fit the experimental data adequately. During and after exposure the following functions may be fitted to the observed C_{Alv} data⁵:

$$C_{Ven} \simeq C_{Alv}(t) = \sum_{i=1}^{p} c(i) (1 - e^{r_i t})$$
 (1)

 $t \leq t_1$

$$C_{Ven} \simeq C_{Alv}(t) = \sum_{i=1}^{p} c(i) (e^{r_i(t-t_i)} - e^{r_it})$$
 (2)

 $t \ge t_1$

where $t_1 = \text{end of exposure}$; p = number of compartments; c(i), $r_i(=\text{neg})$ with i = 1, 2..., p are estimated coefficients and exponents obtained from the fitting. The weighted residual error of the observed data around the fitted curve equals:

$$\sqrt{\frac{1}{n-2p}} \sum_{i=1}^{n} \left(\frac{y_i - \hat{y}_i}{\hat{y}_i} \right)^2 \times 100\%$$
 (3) where y_i = observed value of a sample point (i = 1,

where y_i = observed value of a sample point (i = 1, 2,..n); \hat{y}_i = calculated value by means of the fitted curve with 2p parameters.

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